

OLIGOMYCIN E, A NEW ANTITUMOR
ANTIBIOTIC PRODUCED BY
STREPTOMYCES SP. MCI-2225

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A new oligomycin, oligomycin E (1), and known oligomycins A (2)¹⁾ and B (3)²⁾, were obtained from the culture broth of *Streptomyces* sp. MCI-2225 as cytotoxic compounds against HeLa cells (a uterine carcinoma cell).

This paper reports the fermentation, isolation and structure of oligomycin E (1). Since no report has been made on cytotoxicity of oligomycins [A (2), B (3), C (4)¹⁾, D (rutamycin) (5)^{3,4)} and rutamycin B (6)⁵⁾] against HeLa cells, this paper also deals with the cytotoxicity of 1 and 2~4 in addition to their antibacterial and antifungal activities.

Microorganisms

Streptomyces sp. MCI-2225 was isolated from a soil of Tasmania, Australia. Morphological, cultural and physiological characteristics of this strain will be described in detail elsewhere.

Fermentation

Streptomyces sp. MCI-2225 on an agar slant was inoculated on a seed medium (40 ml) in a 200-ml Erlenmeyer flask. The seed medium (pH 7.0) was composed of maltose syrup 4%, soybean oil 0.3%, soybean meal 2%, Fermedia 1%, Sungrowth L (Sungrowth Co., Osaka) 0.5%, CaCO₃ 0.3%, FeSO₄·7H₂O 0.001%, CoCl₂·6H₂O 0.0001% and NiCl₂·6H₂O

0.0001%. The flasks were incubated on a rotary shaker (210 rpm) for 4 days at 26°C. The seed culture was transferred into the medium described above in a 500-ml Erlenmeyer flask, and the flasks were incubated on a rotary shaker (210 rpm) for 6 days at 28°C.

Biological Assay

Using oligomycin E (1) and oligomycins A, B and C (2~4) obtained commercially (Sigma, St. Louis), the following biological assays were carried out.

For cytotoxicity against HeLa S₃ cells, the method was essentially according to that of MIRABELLI *et al.*⁶⁾. MIC values for antibacterial activity against the bacteria shown in Table 4 were determined by the standard agar dilution method. Antifungal activity against *Pyricularia oryzae* F67-54 (rice blast disease fungus) was evaluated by the method of AKATSUKA *et al.*⁷⁾.

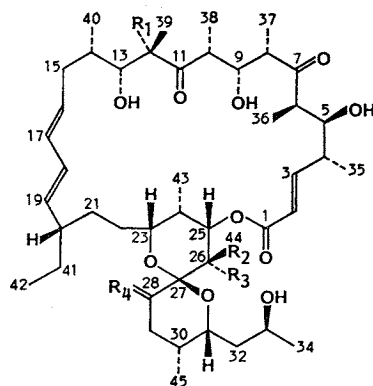
Isolation

The isolation procedure was monitored by cytotoxicity against HeLa cells. The filtered broth (10.8 liters) adjusted to pH 9.0 was extracted with EtOAc (5 liters × 2). A brown paste (7.9 g) from the concentrated extract was chromatographed over Silica gel (Merck, Kieselgel 60, 400 g) eluting with a CHCl₃ - MeOH mixture (95 : 5). The active fractions were combined, and concentrated to give a yellowish oil (5.1 g). Two hundred and fifty mg of the oil was applied on a Pre-sep C18 cartridge (Gasukuro Kogyo Co., Tokyo), and eluted successively with 15 ml each of 20%, 75% and 100% MeOH. This procedure was repeated twenty times to separate all of the yellowish oil. The concentrate of the active 75% MeOH fractions gave a colorless oily residue (424 mg), which was finally purified by HPLC (column: 10.7 × 250 mm packed with Unisil Q C18 (5 μm) (Gasukuro Kogyo Co.); eluent: 80% acetonitrile; flow rate: 5 ml/minute. Components with retention times of 7.9, 9.6 and 12.5 minutes were active, and gave oligomycin E (1) (76 mg), compounds I (3) (73 mg) and II (2) (60 mg), respectively by concentration. Oligomycin E (1) was colorless plates (52 mg) (recrystallized from EtOH). Compounds I (3) (58 mg) and II (2) (48 mg) were colorless prisms when recrystallized from MeOH - water.

Structure

The physico-chemical properties of oligomycin

Fig. 1. Structures of oligomycins.



Oligomycin E (1)	R ₁ =OH,	R ₂ =CH ₃ ,	R ₃ =OH,	R ₄ =O
Oligomycin A (2)	R ₁ =OH,	R ₂ =CH ₃ ,	R ₃ =H,	R ₄ =H ₂
Oligomycin B (3)	R ₁ =OH,	R ₂ =CH ₃ ,	R ₃ =H,	R ₄ =O
Oligomycin C (4)	R ₁ =H,	R ₂ =CH ₃ ,	R ₃ =H,	R ₄ =H ₂
Oligomycin D (5)	R ₁ =OH,	R ₂ =H,	R ₃ =H,	R ₄ =H ₂
Rutamycin B (6)	R ₁ =H,	R ₂ =H,	R ₃ =H,	R ₄ =H ₂

Table 1. Physico-chemical properties of oligomycin E (1).

Nature	Neutral, colorless plates
MP (°C)	120.5 ~ 121.5
[α] _D ²⁵	-49.1° (c 1.05, dioxane)
Formula	C ₄₅ H ₇₂ O ₁₃
<i>Anal</i> Calcd for C ₄₅ H ₇₂ O ₁₃ ·H ₂ O:	C 64.42, H 8.89.
Found:	C 64.46, H 9.12.
FAB-MS (<i>m/z</i>)	821 (M+H) ⁺
UV λ _{max} ^{MeOH} nm (ε)	219 (sh), 224 (29,100), 232 (27,100), 240 (sh)
IR ν _{max} ^{KBr} cm ⁻¹	3456, 1703, 1644

E (1) is summarized in Table 1. The molecular formula of 1 was determined to be C₄₅H₇₂O₁₃ by fast atom bombardment mass spectrometry (FAB-MS) ((M+H)⁺ *m/z* 821) and elemental analysis.

The IR spectrum of 1 indicated the existence of hydroxyl (3456 cm⁻¹) and carbonyl (1703 cm⁻¹) groups. The conjugated diene system was indicated by the IR (1644 cm⁻¹) and UV (λ_{max}^{MeOH} 224 nm) spectra¹¹. These spectral properties suggested 1 to be an oligomycin.

In the ¹H and ¹³C NMR spectra, similar signal patterns were observed between 1 and oligomycin B (3). The comparison of the NMR data between 1 and 3 (see Tables 2 and 3), especially the following signals, led us to determine the structure of 1 as 26-hydroxyoligomycin B (Fig. 1): ¹H NMR, 25-H: d in 1 vs. dd in 3; 26-H: No signal in 1 vs. dq in 3; 44-H₃: s in 1 vs. d in 3; ¹³C NMR, C-26: δ 74.05 in 1 vs. δ 31.27 in 3; C-44: δ 21.52 in 1 vs. δ 11.69 in 3.

The relative stereochemistry of the bicyclic spiroketal moiety of 1 (Fig. 1) was confirmed by the two-dimensional homonuclear nuclear Overhauser effect correlation spectroscopy (NOECOSY) experiment. Cross peaks were observed between: 23-H and 25-H; 23-H and 31-H; 24-H and 25-H; and 25-H and 44-H₃, but no cross peak between 43-H₃ and 44-H₃. Oligomycin E (1) is the first oligomycin having an oxidized C-26.

Compounds I and II were respectively identified as oligomycins B (3) and A (2) by comparison of their physico-chemical properties (mp, [α]_D, IR, UV and NMR) with those of the authentic specimens.

Biological Activities

Biological activities of the oligomycins are listed in Table 4. The order of cytotoxic potency (IC₅₀) against HeLa cells is 2>1=3>4. 1 is effective

Table 2. ¹H NMR data of oligomycins E (1) and B (3) (400 MHz, CDCl₃).

Position	Chemical shift ^a (<i>J</i> , Hz)	
	1	3
2	5.89 (d, 15.4)	5.84 (d, 15.4)
3	7.00 (dd, 15.4, 9.8)	6.69 (dd, 15.4, 10.2)
4	2.42 (ddd, 9.8, 9.8, 6.8)	2.40 (ddd, 10.3, 10.2, 6.8)
5	3.83 (d, 9.8)	3.81 (d, 10.3)
6	2.86 (q, 7.3)	2.76 (q, 6.8)
8	2.72 (qd, 7.3, 3.5)	2.76 (qd, 7.3, 3.0)
9	4.14 (dd, 8.9, 3.5)	3.99 (dd, 8.4, 3.0)
10	3.63 (dq, 8.9, 6.8)	3.65 (dq, 8.4, 6.8)
13	3.90 (s)	3.93 (s)
14	1.91 ^b	1.87 ^b
15	2.01 (ddd, 14.0, 10.9, 10.9), 2.20	2.06 (ddd, 14.0, 11.0, 11.0), 2.15
16	5.48 (ddd, 14.4, 10.9, 3.8)	5.45 (ddd, 13.9, 11.0, 3.7)
17	6.06 (dd, 14.4, 11.0)	6.05 (dd, 13.9, 11.0)
18	5.95 (dd, 14.6, 11.0)	5.94 (dd, 14.6, 11.0)
19	5.23 (dd, 14.6, 9.5)	5.18 (dd, 14.6, 9.5)
20	1.87 ^b	1.84 ^b
21	1.41 ^b , 1.63 ^b	1.34 ^b , 1.61 ^b
22	1.16 ^b , 1.73 ^b	1.08 ^b , 1.63 ^b
23	4.18 (br d, 10.0)	4.05 ^b
24	2.16 ^b	2.15 ^b
25	5.00 (d, 5.9)	4.95 (dd, 11.7, 5.1)
26	—	2.53 (dq, 11.7, 6.8)
29	2.18, 3.18 (dd, 14.2, 5.6)	2.18, 3.04 (dd, 14.7, 5.9)
30	2.30 ^b	2.20 ^b
31	4.53 (br d, 10.2)	4.53 (d, 10.8)
32	1.44 ^b , 1.65 ^b	1.38 ^b , 1.70 (ddd, 13.7, 10.8, 2.2)
33	4.03 ^b	4.08 ^b
34	1.28 (d, 6.8)	1.29 (d, 6.8)
35	1.21 (d, 6.8)	1.19 (d, 6.8)
36	1.04 (d, 7.3)	1.04 (d, 6.8)
37	1.06 (d, 7.3)	1.07 (d, 7.3)
38	1.08 (d, 6.8)	1.09 (d, 6.8)
39	1.14 (s)	1.13 (s)
40	1.01 (d, 7.3)	1.01 (d, 6.8)
41	1.30 ^b , 1.44 ^b	1.28 ^b , 1.40 ^b
42	0.85 (t, 7.3)	0.84 (t, 7.3)
43	1.07 (d, 6.8)	0.86 (d, 6.8)
44	1.29 (s)	0.87 (d, 6.8)
45	0.98 (d, 7.3)	0.96 (d, 7.3)

^a Chemical shift in ppm from TMS.

^b These signals are with unreadable coupling constants because of overlapping with other signals.

—: No signal.

against Gram-positive bacteria, which is different from the other oligomycins. However, all the oligomycins are inactive against Gram-negative bacteria. Although **2**, **3** and **4** show significant antifungal activity (IC₅₀) against *Pyricularia oryzae*, **1** exhibits weak activity (34% inhibition at 100 μg/ml). The order of the antifungal potency is **4** > **2** > **3** > **1**. These activity orders in the

cytotoxicity and antifungal activity suggest that there are some relationships between the activities and the oxidation level at C-12, C-26 and C-28.

No report has been seen on antibacterial activity of the known oligomycins, but C-26 oxidized oligomycin E (**1**) is active against Gram-positive bacteria.

From these activity-oxidation level relation-

Table 3. ^{13}C NMR data of oligomycins E (1) and B (3) (100 MHz, CDCl_3).

Position	1	3	Position	1	3
1	165.44	165.19	24	35.59	35.80
2	122.03	122.47	25	74.20	75.84
3	150.03	148.91	26	74.05	31.27
4	40.43	40.29	27	97.64	100.15
5	73.14	73.01	28	207.89	203.21
6	45.17	46.41	29	45.03	44.11
7	219.77	220.05	30	37.98	37.04
8	46.30	46.03	31	68.01	67.18
9	72.51	72.74	32	41.49	41.70
10	42.11	41.92	33	64.31	64.58
11	220.80	220.36	34	25.11	25.03
12	83.04	83.11	35	17.96	17.94
13	72.51	72.04	36	9.06	9.37
14	33.53	33.66	37	8.64	8.42
15	38.47	38.38	38	14.00	14.02
16	129.77	129.92	39	21.18	21.10
17	132.27	132.23	40	14.44	14.58
18	130.56	130.68	41	28.53	28.71
19	137.38	137.16	42	12.12	12.14
20	45.92	46.15	43	8.61	5.97
21	31.23	31.19	44	21.52	11.69
22	30.37	30.78	45	12.67	12.81
23	70.94	71.15			

Chemical shift in ppm from TMS.

Table 4. Biological activities of oligomycins E (1), A (2), B (3) and C (4).

Organism	1	2	3	4
IC_{50} ($\mu\text{g/ml}$) ^a				
Uterine carcinoma cell				
HeLa S ₃	0.014	0.008	0.015	0.106
MIC ($\mu\text{g/ml}$)				
Bacteria				
<i>Staphylococcus aureus</i> FDA 209PJC 1	6.25	>100	>100	>100
<i>S. aureus</i> Terajima	12.5	>100	>100	>100
<i>S. aureus</i> MS353	12.5	>100	>100	>100
<i>Bacillus subtilis</i> ATCC 6633	100	>100	>100	>100
<i>Micrococcus luteus</i> ATCC 9341	25	>100	>100	>100
<i>Escherichia coli</i> NIHJ JC 2	>100	>100	>100	>100
<i>Klebsiella pneumoniae</i> PCI 602	>100	>100	>100	>100
<i>Salmonella typhimurium</i> IID 971	>100	>100	>100	>100
<i>Serratia marcescens</i> IAM 1184	>100	>100	>100	>100
<i>Pseudomonas aeruginosa</i> IFO 3445	>100	>100	>100	>100
<i>Morganella morganii</i> IFO 3848	>100	>100	>100	>100
<i>Proteus vulgaris</i> HX 19	>100	>100	>100	>100
<i>Providencia rettgeri</i> IFO 3850	>100	>100	>100	>100
<i>Enterobacter aerogenes</i> ATCC 13048	>100	>100	>100	>100
IC_{50} ($\mu\text{g/ml}$) ^b				
Rice blast fungus				
<i>Pyricularia oryzae</i> F67-54	131	3.5	16.9	2.2

^a Concentration causing 50% inhibition of cell growth.^b Concentration causing 50% inhibition of spore germination.

ships, the oxidation level at C-12, C-26 and C-28 is expected to be an important structural factor for various biological activities and to be related to the activity-hydrophilicity relationships.

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